INTENDED USE
The PRONTO® 4 GeneScreen™ kit is a Single Nucleotide Primer Extension Assay, determined by ELISA, intended for the qualitative in vitro detection of the following six mutations: 693C>A and 854A>C in the ASPA gene, 6-bp del/7-bp ins in the BLM gene, IVS4+4 A>T in the FACC gene, 2507+6 T>C and R696P in the IKBKAP gene, in amplified human DNA.

Two assay formats are available:
Option A - simultaneous detection of all six mutations (PRONTO® 4 GeneScreen™ kit).
Option B - separate detection (PRONTO® Canavan or PRONTO® Bloom/Fanconi or PRONTO® FD Screen kits).

For in vitro diagnostic use.

BACKGROUND
Similar to some ethnic populations, the Ashkenazi Jewish population has a higher prevalence of certain genetic disorders. These diseases are inherited in an autosomal recessive pattern. Affected individuals have inherited two copies of the mutated gene, one from each parent.

The following table shows the carrier frequency in the Ashkenazi Jewish population and the detection rate of the conditions tested by the PRONTO® 4GeneScreen™ kit panel.

<table>
<thead>
<tr>
<th>Disease</th>
<th>Mutation</th>
<th>Carrier Frequency</th>
<th>Disease Frequency</th>
<th>Detection Rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Familial Dysautonomia</td>
<td>2507+6 T&gt;C</td>
<td>1/30</td>
<td>1/3,600</td>
<td>98%</td>
</tr>
<tr>
<td></td>
<td>R696P</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Canavan Disease</td>
<td>693C&gt;A 854A&gt;C</td>
<td>1/41</td>
<td>1/6,724</td>
<td>98%</td>
</tr>
<tr>
<td>Fanconi Anemia</td>
<td>IVS4+4 A&gt;T</td>
<td>1/80</td>
<td>1/25,600</td>
<td>99%</td>
</tr>
<tr>
<td>Bloom Syndrome</td>
<td>6-bp del/7-bp ins</td>
<td>1/110</td>
<td>1/48,400</td>
<td>99%</td>
</tr>
</tbody>
</table>
REFERENCES

Bloom Syndrome

Fanconi Anemia C

Canavan Disease

Familial Dysautonomia (FD)

WARNINGS AND PRECAUTIONS

- The Stop Solution contains dilute sulfuric acid (1M), which is an irritant of the eyes and the skin. In case of contact with the eyes, immediately flush them with water. Do not add water to this product. In case of an accident or discomfort consult a physician (if possible, show the bottle label).
- In addition to reagents in this kit, the user may come in contact with other harmful chemicals that are not provided, such as ethidium bromide and EDTA. The appropriate manufacturers’ Material Safety Data Sheets (MSDS) should be consulted prior to the use of these compounds.

ASSAY OVERVIEW

The PRONTO® procedure detects predefined polymorphisms in DNA sequences, using a single nucleotide primer-extension assay. Two steps are carried out prior to the use of this PRONTO® kit:

1. TARGET DNA AMPLIFICATION: The DNA fragments that encompass the tested mutations are amplified. This amplified DNA is the substrate for the primer extension reaction.

2. POST-AMPLIFICATION TREATMENT: The amplified DNA is treated to inactivate free unincorporated nucleotides, so that they will not interfere with the primer extension reaction.

3. PRIMER EXTENSION REACTION: A single nucleotide primer extension reaction is carried out in a 96-well thermoplate. Each well contains a 5'-labeled primer that hybridizes to the tested DNA next to the suspected mutation site, and a single biotinylated nucleotide species (corresponding to mutant or wild type), which complements the nucleotide base at the tested site. Each post-amplification treated sample is tested in two wells per mutation: the first well of each pair tests for the presence of the mutant allele (mut), while the second well tests for the presence of the normal allele (wt). The biotinylated nucleotide will be incorporated in the primer in the course of the reaction or not added, depending on the tested individual’s genotype.

4. DETECTION BY ELISA: The detection of the biotin-labeled extended primer is carried out by an ELISA procedure: The biotin-labeled primers bind to a streptavidin-coated ELISA plate and are detected by a peroxidase-labeled antibody (HRP) directed to the 5’ antigenic moiety of
the primer. A peroxidase reaction takes place in the presence of the substrate - TMB.

6. **INTERPRETATION OF THE RESULTS:** The results are determined either visually (substrate remains clear or turns blue) or colorimetrically (substrate color remains clear or turns yellow) following the addition of the stop solution.

**DISCLAIMER**

- Results obtained using this kit should be confirmed by an alternative method.
- Confirmed results should be used and interpreted only in the context of the overall clinical picture. The manufacturer is not responsible for any clinical decisions that are taken.

The user of this kit should emphasize these points when reporting results to the diagnosing clinician or the genetic counselor.

**MATERIALS PROVIDED**

<table>
<thead>
<tr>
<th>Item</th>
<th>Quantity</th>
<th>Volume/Size</th>
</tr>
</thead>
<tbody>
<tr>
<td>4GeneScreen amplification mix</td>
<td>1 x vial</td>
<td>(0.95µL)</td>
</tr>
<tr>
<td>PRONTO® Buffer 2</td>
<td>1 x bottle</td>
<td>(3 mL)</td>
</tr>
<tr>
<td>Solution C</td>
<td>1 x vial (yellow cap)</td>
<td>(130 µL)</td>
</tr>
<tr>
<td>Solution D</td>
<td>1 x vial (red cap)</td>
<td>(100 µL)</td>
</tr>
<tr>
<td>ColoRed™-Oil</td>
<td>1 x dropper bottle</td>
<td>(13 mL)</td>
</tr>
<tr>
<td>Assay Solution</td>
<td>1 x bottle (green solution)</td>
<td>(100 mL)</td>
</tr>
<tr>
<td>Wash Solution (conc. 20x)</td>
<td>1 x bottle</td>
<td>(100 mL)</td>
</tr>
<tr>
<td>Conjugated HRP</td>
<td>1 x vial</td>
<td>(450 µL)</td>
</tr>
<tr>
<td>TMB- Substrate</td>
<td>1 x bottle</td>
<td>(40 mL)</td>
</tr>
<tr>
<td>Stop Solution (1M H$_2$SO$_4$)</td>
<td>1 x bottle</td>
<td>(30 mL)</td>
</tr>
<tr>
<td>Detection Plates</td>
<td>3 x Streptavidin-coated plates</td>
<td></td>
</tr>
<tr>
<td>PRONTO® 4GeneScreen™</td>
<td>3 x individually pouched plates</td>
<td></td>
</tr>
</tbody>
</table>

**STORAGE AND STABILITY**

- Store at 2-8°C. **Do not freeze.**
- Do not use the kit beyond its expiration date (marked on box label).
- Stability is maintained even when components are re-opened several times.
- Minimize the time reagents spend at room temperature.
- This kit has been calibrated and tested as a unit; do not mix reagents from kits with different lot numbers.

**ADDITIONAL MATERIALS REQUIRED**

- Taq DNA polymerase
- Deionized water (about two liters per kit)
- Thermowell plate or tubes (thin wall) for the post-amplification treatment
- Sterile pipette tips
- Troughs/reagent reservoirs - for use with the detection reagents
- Thermocycler for a 96-well microplate
- Multichannel pipettes (5-50 µL and 50-200 µL)
- Positive displacement pipettes (1-5 µL, 5-50 µL, 50-200 µL & 200-1,000 µL)
- Filtered tips
- ELISA reader with 450 nm filter (optional - 620 nm filter)
- Polaroid camera and color film to record results (optional)
- Automated microtiter plate washer or squirt bottle
- Vortex mixer
- Timer

**ASSAY PROCEDURE**

1. **DNA AMPLIFICATION**

   1. Dispense 2 µL template DNA (from an initial concentration of about 150 ng/µL) to a thermoplate well or tube.
   2. Prepare a Master Mix in a sterile vial, according to the volumes indicated in the table below, plus one spare reaction volume. Add the
Taq DNA polymerase to the Master Mix shortly before dispensing the Mix. Gently mix by pipetting in and out several times.

### PCR Master Mix

<table>
<thead>
<tr>
<th>Solution</th>
<th>Volume for one sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amplification Mix 4GeneScreen™</td>
<td>13.0 µL</td>
</tr>
<tr>
<td>Taq DNA Polymerase (5 u/µL)</td>
<td>0.5 µL</td>
</tr>
</tbody>
</table>

The following Taq DNA polymerases were validated for use with this procedure (lacking 3'→5' exonuclease activity):

- PHARMACIA Cat. # 27-0799
- SIGMA Cat. # D-1806
- ROCHE Cat. # 1-146-165
- PROMEGA Cat. # M-1661
- BIOLINE Cat. # M95801B
- PERKIN ELMER Cat. # M801-0060

1. **Dispense** 13.5 µL Master Mix to each sample.
2. **Add** one drop of ColoRed™ oil to each well. Do not touch the wells with the tip of the oil bottle. Even when using a thermocycler with a hot lid, it is recommended to use oil.
3. **Place** the thermoplate well or tube in a thermocycler previously programmed with the following protocol:

   **Cycling protocol**
   
   1. 94°C 5 minutes
   2. 94°C 30 seconds
   3. 60°C 30 seconds
   4. 72°C 30 seconds
   5. 72°C 5 minutes

4. To verify amplification, **subject** 5 µL of the amplified product to electrophoresis in a 2% agarose gel.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Mutation</th>
<th>Fragment size</th>
</tr>
</thead>
<tbody>
<tr>
<td>ASPA</td>
<td>693 C&gt;A</td>
<td>162 bp</td>
</tr>
<tr>
<td>FACC</td>
<td>IVS4+4A&gt;T</td>
<td>253 bp</td>
</tr>
<tr>
<td>ASPA</td>
<td>854 A&gt;C</td>
<td>285 bp</td>
</tr>
<tr>
<td>BLM</td>
<td>6-bp del/7-bp ins</td>
<td>660 bp</td>
</tr>
<tr>
<td>IKBKAP</td>
<td>2507+6 T&gt;C</td>
<td>802 bp</td>
</tr>
<tr>
<td>IKBKAP</td>
<td>R696P</td>
<td>802 bp</td>
</tr>
</tbody>
</table>

**Limitation of the test:**

Different Taq DNA polymerases and thermocyclers may influence the amplification yield dramatically. It is recommended to use a validated Taq DNA polymerase and a calibrated thermocycler.

2 **POST-AMPLIFICATION TREATMENT**

Only 5 or 10 µL of each amplified DNA sample will be used to carry out this assay.

1. **Prepare** a Post-Amplification Treatment mix shortly before use. Combine in a single test tube the volumes appearing in the following table, multiplied by the number of tested samples, plus one spare volume.
2 Take a PRONTO® Plate out of its pouch. Notice the color at the bottom of the wells. For each mutation tested, use a pink well (mut) and a blue well (wt). Mark the plate with the ID numbers of your test. If you intend to use less than a full plate, you can cut the plate and return the unused portion to the pouch. If you do this, seal the pouch immediately with its desiccant card inside.

3 Primer extension can be carried out in one of two ways:
   - **Option A:** using the combined PRONTO® 4 GeneScreen™ plate for simultaneous detection of six mutations.
   - **Option B:** using separate PRONTO® plates for simultaneous detection of two mutations (Canavan or Bloom/Fanconi or FD).

### PRIMER EXTENSION REACTION

1. **Program** the thermocycler as follows:

<table>
<thead>
<tr>
<th>Cycle</th>
<th>Temperature</th>
<th>Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Start:</td>
<td>94°C</td>
<td>15 seconds</td>
</tr>
<tr>
<td>20 cycles:</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>94°C</td>
<td>30 seconds</td>
</tr>
<tr>
<td></td>
<td>57°C</td>
<td>10 seconds</td>
</tr>
<tr>
<td>End:</td>
<td></td>
<td>Cool down to 25°C (room temperature)</td>
</tr>
</tbody>
</table>

### Post-Amplification Mix

<table>
<thead>
<tr>
<th>Components</th>
<th>Option A (6 mutations)</th>
<th>Option B (2 mutations)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amplified DNA</td>
<td>10.0 µL</td>
<td>5.0 µL</td>
</tr>
<tr>
<td>PRONTO® Buffer 2</td>
<td>90.0 µL</td>
<td>45.0 µL</td>
</tr>
<tr>
<td>Solution C</td>
<td>4.0 µL</td>
<td>2.0 µL</td>
</tr>
<tr>
<td>Solution D</td>
<td>3.0 µL</td>
<td>1.5 µL</td>
</tr>
<tr>
<td>Total Volume</td>
<td>97.0 µL</td>
<td>48.5 µL</td>
</tr>
</tbody>
</table>

2 Mix gently by pipetting this solution in and out five times. Do not vortex.

3 Add 48.5 or 97 µL of the Post-Amplification mix into each well or tube containing 5 or 10 µL of each amplified DNA sample according to the table above. Ensure that the solution you add becomes well mixed with the DNA sample by pipetting.

4 Add one drop of ColoRed™ oil to each tube. Do not touch the tube with the tip of the oil bottle. Even when using a thermocycler with a hot lid, it is essential to use oil.

5 Incubate for 30 minutes at 37°C, then for 10 minutes at 95°C in a thermocycler.

If not used immediately, the treated sample can be kept at 2-8°C for a maximum of four hours.

### Figure 1: Scheme for Dispensing Post-Amplification Treated Samples into the PRONTO® 4 GeneScreen™ Plate.

Recommendation:
Use a new set of tips for each column. Alternatively use the same set of tips, but do not touch the bottom of the wells.
Option B:
Separate test – for Canavan or Bloom-Fanconi or FD Screen (2 mutations):
Starting from the first sample, dispense 8 µl Post-Amplification treated DNA into each one of the four wells in row A as shown in Figure 2. It is possible to transfer up to eight samples simultaneously using a multichannel pipette. Ensure that the solution is at the bottom of each well by inspecting the plate from below. Be sure that the well does not contain air bubbles. Continue with the remaining samples.

Figure 2: Scheme for Dispensing Post-Amplification Treated Samples into Separate PRONTO® Plates.

Familial Dysautonomia ► Bloom/Fanconi ► Canavan ►

Recommendation:
Use a new set of tips for each column. Alternatively use the same set of tips, but do not touch the bottom of the wells.

4 Tilt the plate and add one drop of ColoRed™ Oil to each well. Do not touch the well with the tip of the oil bottle. Even when using a thermocycler with a hot lid, it is essential to use oil.
5 Turn on the thermocycler and start the cycling protocol. Insert the plate when the temperature has reached 90°C.
6 When the thermal cycling is complete, you can proceed to the ELISA assay, or store the reaction products in the refrigerator and carry out the visualization steps within 24 hours.

4 ELISA ASSAY- COLOR DEVELOPMENT

The ELISA assay consists of the following steps:
1. Binding the biotin-labeled extended primer to the Streptavidin-coated plate.
2. Washing away unbound primers.
3. Incubating with the HRP conjugate.
4. Washing away unbound conjugate.
5. Incubating with the TMB Substrate (color development).

The results of this assay can be determined in one of two ways:

a Visually: by monitoring the development of the blue color.
b Colorimetrically: by adding Stop Solution and measuring the absorbance using an ELISA reader at a wavelength of 450 nm (yellow color).

Before proceeding with the ELISA assay make your choice of visual or colorimetric determination of results.

4 PREPARATIONS

- All components used in the detection step should reach room temperature before starting the assay.
- Dilute the 20x Wash Solution to 1x with deionized water. *Diluted solution may be kept at 18-25°C for up to one month.*
- Peel off the plastic cover of the Detection plate. Mark the side of the plate with the kit name and test number.
- Place the PRONTO® plate and the Detection plate side by side, oriented in the same direction (see Fig 3).

4 TRANSFER TO THE DETECTION PLATE
1 Fill a reagent reservoir / trough with the green colored Assay Solution. About 11 mL will be required for a 96-well plate.

2 Add 100 µL of Assay Solution to the bottom of each well in column 1 of the PRONTO® Plate with a multichannel pipette. Gently mix by pipetting in and out 3-4 times.

3 Without changing tips, transfer 100 µL from each well in this column to the first column in the Detection Plate (see Fig. 3). Ensure that the solution at the bottom of all wells of the PRONTO® plate has turned green by inspecting them from below.

**Figure 3:** Transferring the Primer Extension Products from the PRONTO® Plate to Detection Plate.

4 Repeat this procedure, using a new set of tips for each column. It is essential to maintain the order of the samples. 10 µL of oil carried over or 10 µL of the sample left behind will not significantly affect the detection process.

5 Incubate for 10 minutes at room temperature (18-25°C).

### DETECTION BY ELISA

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Visual Detection (Blue color)</th>
<th>Colorimetric Detection (Yellow color)</th>
</tr>
</thead>
<tbody>
<tr>
<td>6. While the incubation of Assay Solution is taking place, dilute the Conjugated HRP in Assay Solution: For every detection plate used (96 well), about 11 mL of diluted conjugate is required. This solution should be freshly prepared each time the test is run.</td>
<td>Dilution: 1:100 (110 µL of conjugated HRP into 11 mL Assay Solution per plate)</td>
<td>Dilution: 1:250 (44 µL of conjugated HRP into 11 mL Assay Solution per plate)</td>
</tr>
<tr>
<td>7. Empty the plate and wash four times with 350 µL 1x Wash Solution. Ensure that the plate is dry after the last wash step.</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>8. Add 100 µL of freshly diluted conjugated HRP to all the wells, with a multichannel pipette.</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>9. Incubate at room temperature.</td>
<td>10 minutes</td>
<td>10 minutes</td>
</tr>
<tr>
<td>10. Wash the plate as in step 7.</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>11. Add 100 µL TMB -Substrate to each well with a multichannel pipette and incubate at room temperature (18-25°C) until blue color appears</td>
<td>15 minutes</td>
<td>15 minutes</td>
</tr>
<tr>
<td>12. Add 100 µL of Stop Solution to each well with a multichannel pipette. The solution will turn yellow immediately.</td>
<td>—</td>
<td>100 µL</td>
</tr>
<tr>
<td>13. The results can be documented using a Polaroid camera with color film (for example – Fuji FP-100C), or by reading the absorbance using an ELISA reader (signal wavelength setting).</td>
<td>Agitate the plate gently and read results at O.D. 620 nm</td>
<td>—</td>
</tr>
<tr>
<td>14. Within two hours read the absorbance using an ELISA reader (single wavelength setting).</td>
<td>—</td>
<td>450 nm</td>
</tr>
</tbody>
</table>
VALIDATION OF THE RESULTS

For Visual Detection:
For every mutation site tested, at least one of the wells should develop a deep blue color. Otherwise, results are invalid for the relevant mutation (Fig. 4).

For Colorimetric Detection:
For every mutation site tested, at least one of the two wells should yield an O.D. > 0.50 reading.

Figure 4: Visual Interpretation of Genotypes

INTERPRETATION OF RESULTS

Important: Heterozygote or homozygote mutant results should be confirmed by retesting. It is recommended to repeat the test with newly extracted DNA.

Criteria for Visual Interpretation
A deep blue color indicates positive signal, while negative signals appear as a clear to pale blue well (see Fig. 5)

Figure 5: Examples of Genotype assignment according to visual inspection of test results

Criteria for Colorimetric Interpretation (O.D 450)
The genotype of each sample is determined according to two criteria:
- The O.D. values of the mut and wt wells.
- The ratio of mut/wt O.D. values.

Calculate the mut/wt ratios by dividing the signal of the mut well by the signal of the wt well.

Identify the correct genotype of each mutation using the table below:

<table>
<thead>
<tr>
<th>Genotype</th>
<th>mut well</th>
<th>wt well</th>
<th>mut/wt ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>O.D. ≤ 0.35</td>
<td>O.D. ≥ 0.5</td>
<td>ratio ≤ 0.5</td>
</tr>
<tr>
<td>Heterozygote</td>
<td>O.D. &gt; 0.5</td>
<td>O.D. &gt; 0.5</td>
<td>0.5 &lt; ratio &lt; 2.0</td>
</tr>
<tr>
<td>Homozygote</td>
<td>O.D. &gt; 0.5</td>
<td>O.D. ≤ 0.35</td>
<td>ratio ≥ 2.0</td>
</tr>
</tbody>
</table>

Samples with values not included in the above table are considered indeterminate and should be retested.
4 GeneScreen™ PROCEDURE SUMMARY

DNA extraction from human whole blood using a validated method.

DNA Amplification:
Volumes per reaction: 2 µL Template DNA + 13.0 µL Amplification Mix + 0.5 µL Taq Polymerase
Cycling protocol: 94°C 5 min → 35 cycles of (94°C 30 sec. / 60°C 30 sec. / 72°C 30 sec.) → 72°C 5 min.

Post Amplification Treatment:
- Volumes for one reaction
  - PRONTO® Buffer 2
    - Solution C
    - Solution D
  - 90.0 µL
  - 4.0 µL
  - 3.0 µL
  - 97.0 µL
- Pipette in and out to mix
- Add the mix into each well containing amplified DNA sample
- Top with ColoRed™ Oil.
- Incubate 30 minutes at 37°C, then 10 minutes at 95°C

Primer Extension Reaction:
- Dispense 8 µL of each Post-Amplification treated DNA into twelve wells (simultaneous Detection) or four wells (separate Detection) of the PRONTO® Plate.
- Top off with ColoRed™ Oil.
- Start the cycling protocol:
  - 94°C 15 sec → 20 cycles of (94°C 30 sec. / 57°C 10 sec.) → Cool
- Insert the PRONTO® Plate in the thermocycler when the temperature has reached 90°C

Detection:
- Add 100 µL Assay Solution to each well in the PRONTO® Plate and mix.
- Transfer 100 µL from each well of the PRONTO® Plate to the respective position in the detection plate. Incubate 10 minutes at RT.
- Empty the wells and wash four times with 350 µL of 1x Wash Solution.

<table>
<thead>
<tr>
<th></th>
<th>Visual Detection</th>
<th>Colorimetric Detection</th>
</tr>
</thead>
<tbody>
<tr>
<td>Add 100 µL of Conjugated HRP to every well and incubate for 10 minutes at room temperature.</td>
<td>Dilution 1:100</td>
<td>Dilution 1:250</td>
</tr>
<tr>
<td>Empty the wells and wash four times with 350 µL of 1x Wash Solution.</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>Add 100 µL of TMB-Substrate to each well and Incubate at RT for:</td>
<td>15 minutes</td>
<td>15 minutes</td>
</tr>
<tr>
<td>Add Stop solution</td>
<td>-</td>
<td>100 µL per well</td>
</tr>
<tr>
<td>Results documentation</td>
<td>Photo or O.D. 620 nm</td>
<td>O.D. 450 nm</td>
</tr>
</tbody>
</table>
For troubleshooting guide, please refer to our website:  
www.prontodiagnostics.com/ts

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COLORED is a trademark of Pronto Diagnostics Ltd.  
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The PRONTO® Technology is covered by US patent 5,710,028, by European patent 0648222 and by corresponding national patents.

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